

# Ground measurements of Solar-induced chlorophyll fluorescence: retrievals methods and practical cases

MaPi Cendrero-Mateo, Sebastian Wineke, Helge Aasen, Javier Pacheco-Labrador, Alasdair McArthur, Alex Damm, Francisco Pinto, Jose Moreno, Luis Alonso, Luis Guanter, Marco Celesti, Micol Rossini, Nues Sabater, Roberto Colombo, Sergio Cogliatti, Uwe Rascher, and Yves Goulas.



COST is supported by the EU Framework Programme Horizon 2020



# Motivation

#### → Update

- State of the art retrieval methods
- State of the art spectrometers which allow the retrieval of F at both O2B and O2A bands

#### REVIEW

#### Remote sensing of solar-induced chlorophyll fluorescence: Review of methods and applications

M. Meroni<sup>a,\*</sup>, M. Rossini<sup>a</sup>, L. Guanter<sup>b</sup>, L. Alonso<sup>c</sup>, U. Rascher<sup>d</sup>, R. Colombo<sup>a</sup>, J. Moreno<sup>c</sup>

\* Remote Sensing of Environmental Dynamics Lab., DISAT, University of Milan-Bicocca, P.zza della Scienza 1, 20126, Milan, Italy

<sup>b</sup> GFZ German Research Centre for Geosciences, Telegrafenberg A17, 14473, Potsdam, Germany

<sup>e</sup> Image Processing Laboratory, LPI, University of Valencia, P.O. Box 22085, Valencia, Spain

<sup>d</sup> Institute of Chemistry and Dynamics of the Geosphere, ICG-3: Phytosphere, Forschungszentrum Jülich, Leo-Brandt-Str, 52425 Jülich, Germany

# Modeling the impact of spectral sensor configurations on the FLD retrieval accuracy of sun-induced chlorophyll fluorescence

Alexander Damm<sup>a,\*</sup>, André Erler<sup>b</sup>, Walter Hillen<sup>c</sup>, Michele Meroni<sup>d</sup>, Michael E. Schaepman<sup>a</sup>, Wout Verhoef<sup>e</sup>, Uwe Rascher<sup>b</sup>

<sup>&</sup>lt;sup>a</sup> Remote Sensing Laboratories, University of Zurich, Winterthurerstrasse 190, CH-8057 Zurich, Switzerland

<sup>&</sup>lt;sup>b</sup> Institute of Bio- and Geosciences, IBG-2: Plant Sciences, Forschungszentrum Jülich, Stetternicher Forst, 52425 Jülich, Germany

<sup>&</sup>lt;sup>c</sup> Aachen University of Applied Sciences, Campus Juelich, Medical Informatics, Heinrich-Mussmann Str. 1, 52428 Jülich, Germany

<sup>&</sup>lt;sup>d</sup> European Commission, DG-JRC, Institute for Environment and Sustainability, Monitoring Agricultural Resources Unit, Via Fermi 2749, 21027 Ispra, Italy

e University of Twente, Faculty of Geo-Information Science and Earth Observation (ITC), P.O. Box 6, 7500 AA Enschede, The Netherlands

# **Under sun-light condition**



# **Under sun-light condition**



Real reflectance ( $\rho$ ) Apparent reflectance ( $\rho_{app}$ )

$$r(\lambda) = \frac{L(\lambda)\pi}{E(\lambda)}$$
$$r_{app}(\lambda) = r(\lambda) + \frac{F(\lambda)\pi}{E(\lambda)}$$

- L = target radiance
- E = solar irradiance
- r = reflectance
- $r_{app}$  = apparent reflectance
- F = Fluorescence

Credits: Meroni et al., 2009 & Jullita 2014

### **Field measurements - Leaf**

![](_page_4_Picture_1.jpeg)

![](_page_4_Figure_2.jpeg)

### **Field measurements – Top of Canopy**

![](_page_5_Picture_1.jpeg)

Credits: Julitta et al. 2016

![](_page_6_Figure_0.jpeg)

Credits: FLEX selection report

#### Solar and atmospheric absorption bands used to retrieve F:

- Hα [645-665 nm]
- O<sub>2</sub>-B [680-700 nm]
- Fe [758.7-758.9]
- O<sub>2</sub>-A [750-770 nm]
- KI [770-770.2 nm]

![](_page_6_Figure_8.jpeg)

### **Sun-Induced Chlorophyll Fluorescence**

![](_page_7_Figure_1.jpeg)

# Brief history of top of canopy sun-induced Cholorphyll fluorescence retrieval methods

1975 - **FLD** 

![](_page_8_Figure_2.jpeg)

# Fraunhofer Line Depth (sFLD)

![](_page_9_Figure_1.jpeg)

#### **ASSUMPTION:**

- Only use two bands one inside and one outside the absorption band
- Fluorescence and reflectance are considered constant inside and outside the absorption band

$$r(\lambda_{in}) \approx r(\lambda_{out})$$
  $F(\lambda_{in}) \approx F(\lambda_{out})$   
1975 – Plascyk, 1975

Credits: addapted from Jullita 2014

# Sun Induced Chlorophyll Fluorescence (SIF) reflectance based Indices

#### **Ratio indices:**

R750/R800 ; R685/R630 ; R680/R630 ; R690/R630; R750/710; R683^2/ (R675\*R691) [Zarco-Tejada et al., 2000]

#### **Derivative index:**

(D688\*D 710)/D697^2 [Zarco-Tejada et al., 2000]

#### In-filling index:

760.59-759.5 [Pérez-Priego et al., 2005]

![](_page_10_Figure_7.jpeg)

![](_page_11_Figure_1.jpeg)

Method	Assumption
3FLD - Maier et al., 2003	Reflectance and fluorescence vary linearly – 1 band inside and 2 outside
cFLD - Gomez-Chova et al., 2006	<b>Reflectance</b> varies according to <b>polynomial functions</b> , <b>fluorescence</b> according to <b>leaf emission</b> – 1 band inside and 2 outside
eFLD - Mazzoni et al., 2007	<b>Reflectance</b> varies according to <b>polynomial functions</b> and <b>fluorescence is</b> <b>determined based on the calculated apparent reflectance</b> – bands inside and 2 outside
iFLD - Alonso et al., 2007 & Damm et al., 2014.	<b>Reflectance</b> varies according to <b>cubic splines functions</b> , and <b>coefficients</b> <b>compensate</b> for using <b>aRFL</b> instead of <b>true RFL</b> – 1 band inside and 2 outside

![](_page_11_Figure_3.jpeg)

![](_page_12_Figure_1.jpeg)

Method	Assumption
3FLD - Maier et al., 2003	Reflectance and fluorescence vary linearly – 1 band inside and 2 outside
cFLD - Gomez-Chova et al., 2006	<b>Reflectance</b> varies according to <b>polynomial functions</b> , <b>fluorescence</b> according to <b>leaf emission</b> – 1 band inside and 2 outside
eFLD - Mazzoni et al., 2007	Reflectance varies according to polynomial functions and fluorescence is determined based on the calculated apparent reflectance
iFLD - Alonso et al., 2007 & Damm et al., 2014.	<b>Reflectance</b> varies according to <b>cubic splines functions</b> , and <b>coefficients</b> <b>compensate</b> for using <b>aRFL</b> instead of <b>true RFL</b> – 1 band inside and 2 outside

 $r(\lambda_{\rm in}) = \alpha_r^* r(\lambda_{\rm out}), \quad F(\lambda_{\rm in}) = \alpha_F F(\lambda_{\rm out})$ 2003 2006 2007 2014

Credits: addapted from Jullita 2014

![](_page_13_Figure_1.jpeg)

Method	Assumption	
3FLD - Maier et al., 2003	Reflectance and fluorescence vary linearly – 1 band inside and 2 outside	
cFLD - Gomez-Chova et al., 2006	<b>Reflectance</b> varies according to <b>polynomial functions</b> , <b>fluorescence</b> according to <b>leaf emission</b> – 1 band inside and 2 outside	
eFLD - Mazzoni et al., 2007	Reflectance varies according to polynomial functions and fluorescence is determined based on the calculated apparent reflectance	
iFLD - Alonso et al., 2007 & Damm et al., 2014.	<b>Reflectance</b> varies according to <b>cubic splines functions</b> , and <b>coefficients compensate</b> for using <b>aRFL</b> instead of <b>true RFL</b>	

$$F(\lambda) = L(\lambda) - FIT(r^*(\lambda))E(\lambda) / \pi$$
Credits: addapted from Jullita 2014

![](_page_14_Figure_1.jpeg)

Method	Assumption					
3FLD - Maier et al., 2003	Reflectance and fluorescence vary linearly – 1 band inside and 2 outside					
cFLD - Gomez-Chova et al., 2006	<b>Reflectance</b> varies according to <b>polynomial functions</b> , <b>fluorescence</b> according to <b>leaf emission</b> – 1 band inside and 2 outside					
eFLD - Mazzoni et al., 2007	Reflectance varies according to polynomial functions and fluorescence is determined based on the calculated apparent reflectance					
iFLD - Alonso et al., 2007 & Damm et al., 2014.	<b>Reflectance</b> varies according to <b>cubic splines functions</b> , and <b>coefficients</b> <b>compensate</b> for using <b>aRFL</b> instead of <b>true RFL</b>					
	$\alpha_F^* \approx \frac{E(\lambda_{\text{out}})}{\tilde{E}(\lambda_{\text{in}})} \alpha_r^*$					

Credits: addapted from Jullita 2014

2003 2006 2007

2014

![](_page_15_Figure_0.jpeg)

# **Peak Height Method**

![](_page_16_Figure_1.jpeg)

# Spectral Fitting Methods (SFM)

![](_page_17_Figure_1.jpeg)

Credits: addapted from Jullita 2014

# **Spectral Fitting Methods (SFM)**

Cost function optimization:

$$min\sum(L-L_{mod})^2$$

2006

![](_page_18_Figure_3.jpeg)

# From theory to practice

How to apply these methods to measured data?

Not always easy to translate the method into an algorithm

**Research questions:** 

- 1. Sensitivity to Spectral resolution and SNR
- 2. Sensitivity to Spectral shift
- 3. Wavelength selection in and out the absorption band

![](_page_19_Figure_7.jpeg)

# Model data

Table 1 - Spectrometer characteristics:

Spectrometer	Range [nm]	SSI [nm]	FWHM [nm]	SNR
HR4000	650-840	0.05	0.28	300:1
ΜΑΥΑ	650-803	0.08	0.44	450:1
QEPRO	651-803	0.13	0.38	1100:1
ASD	350-1000	1.4	3	4000

\*adapted from: Celesti et al. 2016. Comparison of sun-induced chlorophyll fluorescence estimates from commercial spectrometers: an optimal setup for field measurements and aerial product validation

#### Spectra

- The 1600 spectra per spectral library comprise the following:
   0 16 vegetation types \_\_\_\_\_\_
  - o each vegetation type is simulated 100 times
  - the first spectra per set of 100 spectra is without noise
  - the following 99 spectra are replicates but with noise

Spectra	Name	LAI	LIDF	CAB	f-eff	SIF685	SIF760
1	L1_E_20_2.txt	1	Erectophile	20	2	0.7617	0.3383
2	L1_E_20_4.txt	1	Erectophile	20	4	1.5234	0.6766
3	L1_E_80_2.txt	1	Erectophile	80	2	0.5589	0.402
4	L1_E_80_4.txt	1	Erectophile	80	4	1.1178	0.8039
► 5	L1_P_20_2.txt	1	Planophile	20	2	0.6378	0.6378
6	L1_P_20_4.txt	1	Planophile	20	4	2.9226	1.2756
7	L1_P_80_2.txt	1	Planophile	80	2	1.1293	0.76
8	L1_P_80_4.txt	1	Planophile	80	4	1.52	1.52
9	L4_E_20_2.txt	4	Erectophile	20	2	1.167	0.5164
10	L4_E_20_4.txt	4	Erectophile	20	4	2.3339	1.0329
11	L4_E_80_2.txt	4	Erectophile	80	2	0.8428	0.595
12	L4_E_80_4.txt	4	Erectophile	80	4	1.6856	1.1899
13	L4_P_20_2.txt	4	Planophile	20	2	1.921	0.8378
14	L4_P_20_4.txt	4	Planophile	20	4	3.842	1.6757
15	L4_P_80_2.txt	4	Planophile	80	2	1.4659	0.9826
16	L4_P_80_4.txt	4	Planophile	80	4	2.9318	1.9652

# 1. Sensitivity to Spectral resolution and SNR

![](_page_21_Figure_1.jpeg)

#### **Retrieval methods**

- sFLD, 3FLD, iFLD (done!)
- SFM, Peak Height (next...)

![](_page_21_Picture_5.jpeg)

#### O<sub>2</sub>-B band results (F687) – all 16 vegetation types WITHOUT noise

![](_page_22_Figure_1.jpeg)

#### O<sub>2</sub>-A band results (F760) – all 16 vegetation types WITHOUT noise

![](_page_23_Figure_1.jpeg)

#### $O_2$ -A band results (F760) – all 16 vegetation types WITH noise

![](_page_24_Figure_1.jpeg)

- → Plot Average standard deviation of retrieved F
- Low SNR levels (10-200) led to a high fluctuation of the measured radiance signals and consequently to high F retrieval error.
- sFLD insensitive to noise reached stable signal SNR ~ 200
- 3FLD reached stable signal SNR ~ 1000
- iFLD sensitive to noise no stable error value was fond even SNR ~ 10000
- Next similar analysis with O<sub>2</sub>B band and peak height and SFM methods.

# 2. Sensitivity to Spectral shift

![](_page_25_Figure_1.jpeg)

![](_page_25_Figure_2.jpeg)

- → Spectral miscalibration of the up looking and down looking channel.
- → Plot Average absolute difference between reference F and retrieved F
- sFLD most affected by spectral shift
- Lower spectral resolution higher the influence of the spectral shift in F (spectral shift = 0.5 spectral resolution)

# 3. FLD like approaches: Wavelength selection in and out the absorption band

#### Recommendations

#### Channel inside:

• set to the position of the minimal in each of the absorption bands.

#### Channel outside:

- the irradiance should be relatively steady around the channels (which means the channels are on the "shoulders" of the absorption valleys)
- the channels should be as near to the inside channels as possible.

![](_page_26_Figure_7.jpeg)

O<sub>2</sub>-A band results (F760) – iFLD

![](_page_27_Figure_1.jpeg)

- The error in the FLD-based methods are mainly caused by the F and reflectance different between the bands inside and outside the absorption bands.
- The RMSE increase when the distance between in the inside and outside absorption band increase (it is lower spectral resolution higher RMSE)

# **Conclusions and future work**

#### General conclusions (O<sub>2</sub>B and O<sub>2</sub>A band)

- When no noise is taking into account ASD-Low spectral resolution worst results and HR400-High spectral resolution best results.
- When noise is taking into account QEPRO lower spectral resolution compare to HR400 but better SNR 1100:1 best performance.
- FLD-bases method band selection,
  - Inside the absorption band lower point
  - Outside the absorption band maximum points ~ avoid absorption valleys
  - Selection of bands ~ reduce distance between inside and out side bands

#### O<sub>2</sub>B band (F680)

- Non of the FLD-based method provide an accurate estimation of F680
- Future we need to evaluate peak height and SFM performance

#### $O_2A$ band (F760) ~ Damm et al 2010.

#### sFLD

- Strongly overestimate F
- Sensitive to small signal variations
- 1- Spectral resolution
- 2- then spectral shift and SNR

#### 3FLD

- Best compromise between robustness and accuracy
- 1- SNR
- 2- then spectral resolution and spectral shift

#### iFLD

- Best performance but highly influence by noise
- 1 SNR
- 2 then spectral resolution and spectral shift

#### **O**<sub>2</sub>B band (F680)

- Non of the FLD-based method provide an accurate estimation of F680 ٠
- Future we need to evaluate peak height and SFM performance •

#### O<sub>2</sub>A band (F760) ~ Damm et al 2010.

#### sFLD

- Strongly overestimate F
- Sensitive to small signal variations
- 1- Spectral resolution
- 2- then spectral shift and SNR •

#### 3FLD

- Future-wereed to evaluate peak height and Best compromise between robustness and accuracy
- **1- SNR** ٠
- 2- then spectral resolution and spectral shift •

#### iFLD

- Best performance but highly influence by noise
- 1 SNR
- 2 then spectral resolution and spectral shift

![](_page_31_Picture_0.jpeg)

![](_page_31_Picture_1.jpeg)

# Thanks for your attention!!

![](_page_31_Picture_3.jpeg)